DOI: http://dx.doi.org/10.18782/2320-7051.5947

ISSN: 2320 – 7051 *Int. J. Pure App. Biosci.* **5 (6):** 770-777 (2017)



Research Article



CRH Gene Polymorphism Associated with Milk and Its Constituents in Karan Fries Cattle

Jyoti Beniwal, Anupama Mukherjee^{*}, Alok Kumar Yadav and Shabahat Mumtaz

Division of Animal genetics and Breeding, ICAR-National Dairy Research Institute, Karnal, Haryana, India *Corresponding Author E-mail: writetoanupama@gmail.com Received: 27.10.2017 | Revised: 29.11.2017 | Accepted: 6.12.2017

ABSTRACT

The aim of this study was to estimate the relations between the CRH-A787G polymorphism and milk production traits (first lactation 305 days milk yield, first lactation total milk yield, first lactation 305 days protein and first lactation fat yield, as well as first lactation 305 days lactose and first lactation 305 days SNF yield) in 60 Karan Fries cows. The genotype and allele frequencies were estimated and they were as follows: AG - 0.63; AA - 0.37; A - 0.69; G - 0.31. Statistical analysis revealed that studied polymorphism significantly affected the first lactation 305 days milk yield, total milk yield, 305 days fat and SNF yield ($P \le 0.01$). The results indicate that selection for the CRH-A787G, AG animals might contribute to increase the value of these traits in Karan Fries cattle. However, further studies are necessary to verify the results of our study.

Key words: Karan Fries cattle, Milk Yield, Milk Constituents traits.

INTRODUCTION

Agriculture is the main stay of Indian economy as agriculture and allied sectors contribute nearly 14 % of Gross Domestic Production. Livestock sector alone contributes nearly 25.6% of value of output in Agriculture, Fishing and Forestry sector¹. About 58% of population is engaged in agriculture and rearing of livestock in the country. As per All India Livestock Census, 2012. India has 190.90 million (37.28%) cattle and 108.7 million (21.23%) buffaloes. Out of total cattle population, about 39.73 million are crossbred in our country. The decade wise trend in livestock population (1997 to 2012) shows a distinct shift in composition of dairy animal stock in favor of buffaloes and crossbred cattle, as their numbers increased by 3.19% and 20.18%, respectively, while that of indigenous cattle declined by 8.94% (19th Livestock Census). Milk productivity in the country remains one of the lowest as compared to many leading countries of the world. In India, average milk productivity of crossbred cows, indigenous cows and buffaloes is about 7.02, 2.36 and 4.8 kg/day, respectively (Ministry of Agriculture, GOI, 2013-14). India has always been 100% self sufficient in milk, with total imports/exports of about 0.3 million tonnes per annum and thus it may be considered as almost unconnected with the world dairy market.

Cite this article: Beniwal, J., Mukherjee, A., Yadav, A.K. and Mumtaz, S., CRH Gene Polymorphism Associated with Milk and Its Constituents in Karan Fries Cattle, *Int. J. Pure App. Biosci.* **5(6):** 770-777 (2017). doi: http://dx.doi.org/10.18782/2320-7051.5947

ISSN: 2320 - 7051

The economic survey 2011 analyzed the dairy situation in India, considering that the requirement of milk in 2021 to 2022 is expected to be 180 million tones as against the current level of milk production of 137.7 million tonnes (BAHS,2015).

One of the major constrains in genetic progress of dairy animal is that milk production traits are controlled by several genes which are expressed later in life. Recent advances of molecular genetics in identification of QTLs affecting production traits of domestic animals have opened new vistas for genetic improvement of economic traits. . It is noteworthy that OTLs for milk performance traits have been mapped to all the bovine autosomes, mostly to autosome 6, 14 and 20⁹. Moreover, mutations in candidate genes for the above-mentioned traits are identified and analyzed^{7,12,13}. The positional and functional candidate gene approach have been applied to different genes in dairy animals .GH, DGAT1, SCD1, PRL, STAT5A, OLR1, LEP, LGB, ABCG2, CSN3 are some of the important candidate genes for milk production traits. First positional clone of QTL in cattle was done for DGAT1 that is associated with fat yield⁵.

Corticotropin-releasing hormone called also corticoliberin (CRH), or corticotropin-releasing factor (CRF), is a 41amino acid peptide deriving from a 191-amino acid precursor. It is synthesized mainly in the hypothalamus, but also in other brain areas. The highest level of the CRH gene expression was found in the hypothalamus, but this gene is also expressed in many other places, such as placenta, uterus, ovaries, testes, liver, stomach, skin, immune system. CRH functions as a neuropeptide hormone participating, among others, in the stress response in vertebrates. Corticotropin-releasing hormone is also involved in controlling the energy balance of an organism, and thus can affect body weight. In addition, it participates in modulating immune and reproductive systems¹⁵. The gene encoding corticoliberin has been mapped to the bovine chromosome 14 2 , where QTLs for the postnatal growth have also been identified.

Therefore, it has been considered a candidate gene for growth traits in cattle³. Taking into account the position of this gene, in the proximity of the QTL for milk performance traits, it can be considered a positional candidate gene for the abovementioned traits. Within the bovine CRH gene, several single nucleotide polymorphic (SNP) sites have been identified: C22G causing substitution of amino acids in the signal sequence, A145G and C240G, leading to changes of amino acids in a propeptide³ as well as two SNPs in exon 2 14 . The undertaken study aimed at determining the frequencies of alleles and genotypes with regard to the polymorphism in the gene corticotropin-releasing encoding hormone (CRH-A787G) and establishing association between the genotypes and some milk production traits of Karan Fries cows.

MATERIALAND METHODS

The study involved a total of 60 Karan Fries cows from the herd kept in the Livestock Research Center (LRC) of ICAR-National Dairy Research Institute, Karnal (Haryana), India. DNA isolation was done by collecting 10 ml blood aseptically by jugular vein puncture in a sterile Vacutainer (Beckton-Dickinson vacutainer containing 0.5% EDTA solution (10µl/ml of blood). The samples were transported to the laboratory in an ice box and stored at -20[°]C till further processing for DNA isolation. DNA Extraction from blood was Phenol chloroform method ,as done by described by Sambrook and Russel with minor modifications was used for DNA isolation from blood of Karan Fries Cattle .

In investigated SNP adenine to guanine at position 787 in exon 1 – *CRH*-*A787G* has been observed. DNA sequencing (First Base Laboratories) was used for genotyping and the sequenced data was analysed by BLAST (version 1.2.0) and Clustal W. A set of forward and reverse gene – specific oligonucleotide primer was designed to amplify CRH gene using Primer 3 software (http://www.primer3.ut.ee) (Untergrasser *et al.*, 2012) and gene sequence available at NCBI database (http://www.ncbi.nlm.nih.gov) . The

ISSN: 2320 - 7051

Beniwal *et al*

primer designed was checked for specificity by BLAST (version 1.2.0). Primer was designed and synthesized from Sigma Aldrich Chemicals Pvt. Ltd (USA). The sequence of primer from 5' to 3' end and its length is 353 bp respectively was designed.

F 5'-CTGCGTGGTTTCTGAAGAGG-3' R 3'CGCGTTCACACACAACAC-5'

The gene fragment of 353 bp was amplified. The amplification reaction thermal program was as follows: initial denaturation at 95°C for 3 min, followed by 30 cycles: 94°C for 30 s (denaturation), 55°C for 30 s (primer hybridization), 72°C for 30 sec (product synthesis), and final extension at 72°C for 10 min. After PCR amplification, the PCR product was checked on 0.8% agarose gel to verify the amplification of target region.

At the end of run, power supply was turned off and the gel was carefully removed from the chamber and placed on trans illuminator for visualization of Gel and viewed under UV light and photographs were taken with the help of gel documentation system. A single sharp band indicated proper amplification of the target DNA.

The amplified PCR products from the set of primer was sent for DNA sequencing to st Base sequencing INT (Singapore) for purification and custom sequencing from both ends (5'and 3' ends). DNA sequence was aligned with corresponding reference sequence using Clustal W multiple sequence alignment program for DNA

(www.ebi.ac.uk/tools/msa/clustalw2)¹⁰ to identify SNPs.

The data on various economic traits during last 8 years (2007-2014) were collected from the history-cum pedigree sheet and milk constituents register maintained under AGB division and LPM Section, NDRI, Karnal The following model will be used for all lactation (Haryana), India respectively .The following information was collected: Animal number, Date of birth, Sire number, Dam number, Lactation number. Traits Date of calving, considered (Parity, Period of calving, Season of calving, first lactation 305 days milk yield, first lactation total lactation milk yield, first lactation test day fat yield, fat content (%), first lactation test day protein yield, protein content(%), first lactation test day lactose yield, first lactation test day SNF yield), traits generated (first lactation 305 day fat yield, first lactation 305 day protein yield, first lactation 305 day lactose yield, first lactation 305 day SNF yield, age at first calving).

As a part of the characterization of the genetic structure of examined herd, the frequencies

of the CRH-A787G genotypes and alleles were estimated and 2 out of the 3 possible genotypes were identified. The next stage was the analysis of the association between genotypes and the values of the following milk production traits in the first lactation: first lactation 305 days milk yield, first lactation total lactation milk yield, first lactation 305 day fat yield, first lactation 305 day protein yield, first lactation 305 day lactose yield, first lactation 305 day SNF yield (kg). The production records of the animals were collected from the breeding documentation carried out for herd as a part of milk recording. Statistical analysis of the data was done by Least squares analysis of variance for unequal and non-orthogonal data using the technique described by Harvey⁶ will be used to study effect of non-genetic factors. The model will be used with assumptions that different components being fitted into the model are linear, independent and additive.

The following model will be used for all lactation traits:

```
\begin{array}{lll} \mathbf{Y}_{ijkm} &= \boldsymbol{\mu} + \mathbf{P}_i + \mathbf{S}_j + \mathbf{A}_k + \mathbf{e}_{ijkm} \\ \text{Where,} \\ \mathbf{Y}_{ijkm} &= m^{th} \text{ observation in } i^{th} \text{ period }, j^{th} \text{ season, } k^{th} \text{ age at first calving } \\ \boldsymbol{\mu} &= \text{Overall mean} \\ \mathbf{P}_i &= \text{Effect of } i^{th} \text{ period } (i = 1 \text{ to } 3) \\ \mathbf{S} &= \text{Effect of } j^{th} \text{ season } (j = 1 \text{ to } 4) \\ \mathbf{A}_k &= \text{Effect of } k^{th} \text{ age st first calving } (k = 1 \text{ to } 5) \\ \mathbf{e}_{ijklm} &= \text{Random error} \sim \text{NID} (0, \sigma^2_e) \end{array}
```

Int. J. Pure App. Biosci. 5 (6): 770-777 (2017)

ISSN: 2320 - 7051

For association Analysis mixed effect model of multifactor analysis of variance (ANOVA), using the GLM (General Linear Model) procedure was used which was based on the available records pertaining to milk yield and its constituents on Karan Fries cattle

Allelic and genotypic frequencies and their accordance with or deviation from Hardy-Weinberg law will be studied using POPGENE software package. Effect of SNP, found significantly contributing towards milk yield and constituents, will be analyzed as under:

 $Y_{ij} = \mu + G_i + e_{ij}$

Where,

 μ = Overall mean

 $G_i =$ Fixed effect of ith genotype

 e_{ij} = Random error ~ NID (0, σ_e^2)

RESULTS AND DISCUSSION

DNA sequencing analysis of the amplified fragment of the CRH gene enabled distinguishing two genotypes: AG and GG, determined by the presence of alleles and genotype amounted to: AA - 0.37; AG - 0.63; A - 0.69; G - 0.39. In the Table 01 and 02 shows effect of genetic and non genetic factors on milk production traits in the First lactation in Karan Fries cattle. The results of the study showed statistically significant differences in the milk production traits for different genotypes and period of calving (First lactation 305 days milk yield, First lactation total lactation milk yield, First lactation 305 day fat yield, First lactation 305 day SNF yield for genotypes, First lactation 305 day protein yield, First lactation 305 day lactose yield). Cows with the AGgenotype were characterized by significantly higher values for the above-mentioned traits compared with the AA cows.

maintained at National Dairy Research Institute, Karnal (Haryana), India an attempt will be made to find the association of different allelic variants of CRH gene with the milk and its constituents.

The approach aimed at identifying mutations in candidate genes facilitates discovering and locating major-effect genes (in particular functional mutations within these genes) for quantitative traits. The candidate genes strategy is used for various genes whose products may affect production traits of cattle. In dairy cattle, genes encoding, among others, milk proteins (e.g. αS1-casein), enzymes involved in the metabolism of fatty acids (e.g. SCD, DGAT1), some hormones (e.g. leptin, growth hormone) or transcription factors (e.g. PIT-1) are intensively analyzed. Significant associations between polymorphisms in these genes and milk performance traits have been found^{4,8,11,17}.

The results of the conducted study indicate that the polymorphism within the corticotropin- releasing hormone gene (CRH-A787G) is associated with an increased first lactation 305 days milk yield, first lactation total lactation milk yield, first lactation 305 day fat yield, first lactation 305 day SNF yield in Karan Fries cattle. However, due to the lack of accessible literature data that would concern the analysis of the association between the above-mentioned polymorphism and milk production traits of cattle, the verification of the obtained results is impossible. The research on the association between the CRH gene polymorphism and production traits in cattle has only been conducted in beef cattle herds. Three polymorphisms within the CRH gene have been analyzed in association with meat related traits³.

Copyright © Nov.-Dec., 2017; IJPAB



Lane1-10 : PCR product (353 bp)

Lane M : 100 bp ladder



Fig 02: Chromatogram showing change of A787G in Karan Fries cattle

Beniwal et alInt. J. Pure App. Biosci. 5 (6): 770-777 (2017)ISSN: 2320 - 7051Table 1: Least square mean and standard error of milk production traits in Karan Fries cattle

FACTORS	N	FL305DMY	FLTMY	FL305DFY	FL305DPY	FL305DLY	FL305DSNFY		
Season of calving									
Dec - Mar	27	3590.99 ± 97.31	5298.34 ± 426.87	155.16 ± 4.05	86.94 ± 10.21	132.13 ± 15.22	$299.35 \pm \ 9.19$		
April - June	21	3432.77 ± 106.81	4479.86 ± 468.56	148.08 ± 4.45	105.51 ± 15.80	161.01 ± 23.55	310.26 ± 14.23		
July - Sept	8	3581.17 ± 165.30	4700.53 ± 725.17	155.10 ± 6.89	109.46 ± 20.72	164.51 ± 30.89	319.56 ± 18.66		
Oct - Nov	4	3663.04 ± 216.76	5189.03 ± 950.91	161.67 ± 9.03	130.16 ± 14.08^{a}	196.68 ± 20.98 ^a	329.61 ± 12.68^{a}		
Period of calving									
2007-10	12	3808.74 ± 147.27^{a}	4984.94 ± 646.06	166.48 ± 6.14^{a}	86.61 ± 10.16^{b}	129.61 ± 15.15^{b}	309.14 ± 9.15^{ab}		
2011-12	33	3541.11 ± 106.35 ^{ac}	4686.62 ± 466.53	152.17 ± 4.43^{b}	79.79 ± 11.49 ^b	122.27 ± 17.14^{b}	292.66 ± 10.35^{b}		
2013-14	15	3351.13 ± 120.27 ^{bc}	5079.02 ± 527.62	146.37 ± 5.01^{b}	111.67 ± 14.48	168.94 ± 23.55	320.18 ± 13.04		
Group of age at first calving									
<947	9	3674.99 ± 151.51	5503.20 ± 664.65	161.91 ± 6.31	111.77 ± 13.05	167.86 ± 19.46	317.29 ± 11.76		
948-1065	12	3647.15 ± 136.58	4489.99 ± 599.15	156.20 ± 5.69	86.23 ± 12.50	130.61 ± 18.64	297.47 ± 11.26		
1066-1183	16	3416.22 ± 130.84	4412.28 ± 573.98	147.74 ± 5.45	81.18 ± 11.43	123.54 ± 17.04	305.14 ± 10.29		
1184-1301	15	3503.53 ± 119.57	5168.87 ± 524.56	153.20 ± 4.98	103.39 ± 17.39	156.64 ± 25.92	312.26 ± 15.66		
>1302	8	3593.07 ± 181.91	5006.95 ± 798.03	155.97 ± 7.58	98.85 ± 7.89	149.52 ± 11.77	310.47 ± 7.11		

The mean values with different superscript alphabet indicate highly significant difference (p<0.01) among themselves

Table 2: Generalised linear model (GLM) of milk production traits in cows with different CRH-A787G genotypes

8									
Trait	SNP/Genotype	Ν	Breeding Value ± SE	CV%					
FL305DMY									
	AA	22	$3129.10^{a} \pm 88.0$	13.19					
	AG	38	$4237.80^{b} \pm 77.9$	11.33					
FLTMY									
	AA	22	$4191^{a} \pm 235$	26.33					
	AG	38	$6279^{b} \pm 325$	31.93					
FL305DFY									
	AA	22	$13.47^{a} \pm 3.89$	13.53					
	AG	38	$183.04^{b} \pm 3.31$	11.15					
FL305DPY									
	AA	22	77.70 ± 7.30	44.08					
	AG	38	98.70 ± 7.74	48.36					
FL305DLY									
	AA	22	117.40 ± 10.92	43.63					
	AG	38	148.52 ± 11.60	48.14					
FL305DSNFY									
	AA	22	$273.07^{a} \pm 7.50$	12.89					
	AG	38	$369.20^{b} \pm 6.63$	11.06					

N – Number of individuals with the genotype

CONCLUSION

In order to use the obtained results for the improvement of First lactation 305 days milk yield, First lactation total lactation milk yield, First lactation 305 days fat yield, First lactation 305 days SNF yield in Karan Fries cattle, the inclusion of individuals with the *AG* genotype in the breeding work could be considered. However, due to the absence of the individuals with the *AA* genotype in the examined herd, further research involving much larger herd as well as herds of other dairy cattle breeds is necessary. It would allow to verify the obtained results prior to their potential application in selection programs of dairy cattle.

REFERENCES

- 19th Livestock Census All India Report, DAHD, & F., Ministry of Animal Husbandry, Dairying and Fisheries , Ministry of Agriculture , Government of India , (2013-14). (2012). (www.dahd.nic.in)
- Barendse, W., Vaiman, D., Kemp, S.J., Sugimoto, Y., Armitage, S.M., Amediumdensity genetic linkage map of the bovine genome. *Mamm. Genet.* 8: 21–28 (1997).
- F.C., 3. Buchanan, Thue, T.D., Yu, P., Winkelman-Sim, D.C., Single nucleotide polymorphisms the in corticotrophin-releasing hormone and proopiomelanocortin genes are associated with growth and carcass yield in beef cattle. Anim. Genet. 36: 127-131 (2005).
- Citek, J., Rehout, V., Hradecka, E., Vecerek, L., Panicke, L., The breeding values of German Holstein sires and the *DGAT1* polymorphism. *Arch. Tierz.* 50: 136–146 (2007).
- Grisart, B., Coppieters, W., Farnir, F., Karim, L., Ford, C., Berzi, P., Cambisano, N., Mni, M., Reid, S., Simon, P., Spelman, R., Georges, M. and Snell, R., Positional candidate cloning of a QTL in dairy cattle : identification of a missense mutation in the bovine DGAT1 gene with major effect on milk yield and composition. *Genome Res.* 12: 222-231 (2002).

- Harvey, W.R., User's Guide for LSMLMW and MIXMDL, Mixed Model Least Squares and Maximum Likehood Computer Program. PC-2 version. The Ohio State University, Columbus, USA (1990).
- Javanmard, A., Khaledi, K., Asadzadeh, N., Solimanifarjam, A.R., Detection of Polymorphisms in the Bovine Leptin (LEP) Gene: Association of a Single Nucleotide Polymorphism With Breeding Value of Milk Traits in Iranian Holstein Cattle. J. Mol. Genet. 2: 10–14 (2010).
- Kaminski, S., Rymkiewicz-Schymczyk, J., Wojcik, E., Rusc, A., Associations between bovine milk protein genotype and haplotypes and the breeding value of Polish Black-and-White bulls. *J. Anim. Feed Sci.* 11: 205–221 (2002).
- Khatkar, M.S., Thomson, P.C., Tammen, I., Raadsma, H.W., Quantitative trait loci mapping in dairy cattle: review and metaanalysis. *Genet. Sel. Evol.* 36: 163–190 (2004).
- Larkin, M.A., Blackshields, G., Brown, N.P., Chenna, R., McGettigan, P.A., McWilliam, H., Valentin, F. and Wallace, I.M., ClustalW and ClustalX version 2.0. *Bioinformatics*, 23: 2947–2948 (2007).
- Liefers, S.C., Te Pas, M.F.W., Veerkamp, R.F., van der Lende, T., Associations between leptin gene polymorphisms and production, live weight, energy balance, feed intake, and fertility in holstein heifers. *J. Dairy Sci.* 85: 1633–1638 (2002).
- Ordovas, L., Zaragoza, P., Altarriba, J., Rodellar, C., Identification of 14 new single nucleotide polymorphisms in the bovine SLC27A1 gene and evaluation of their association with milk fat content. *J. Dairy Res.* **75:** 129–134 (2008).
- Pariset, L., Caroli, A., Chessa, S., Fontanesi, L., Russo, V., Bagnato, A., Schiavini, F., Samoré, A.B., Feligini, M., Bonizzi, I., Vicario, D., Rossoni, A., Sangalli, S., Marino, R., Perini, D., Nicolazzi, E.L., Macciotta, N.P.P., Ajmone-Marsan, P., Assessment of 29

Copyright © Nov.-Dec., 2017; IJPAB

candidate genes for milk traits in Italian dairy cattle. *J. Dairy Res.* **75:** 129–134 (2008).

- 14. Sherman, E.L., Nkrumah, J.D., Murdoch, B.M., Li, C., Wang, Z., Fu, A., Moore S.S., Polymorphisms and haplotypes in the bovine neuropeptideY, growth hormone receptor, ghrelin, insulin like growth Factor 2, and uncoupling proteins 2 and 3 genes and their associations with measures of growth, performance, feed efficiency, and carcass merit in beef cattle. *J. Anim. Sci.* 86: 1–16 (2008).
- Slominski, A., Wortsman, J., Luger, T., Paus, R., Solomon, S., Corticotropin releasing hormone and proopiomelanocortin involvement in the

cutaneous response to stress. *Physiol. Rev.* **80:** 879–1020 (2000).

- Untergasser, A., Cutcutache, I., Koressaar, T., Ye, J., Faircloth, B.C., Remm, M., Rozen, S.G., Primer3- new capabilities and interfaces. *Nucleic Acids Res.* 40(15): 115 (2012).
- Zwierzchowski, L., Krzyżewski, J., Strzałkowska, N., Siadkowska, E., Ryniewicz, Z., Effects of polymorphism of growth hormone (GH), Pit-1, and leptin (LEP) genes, cow's age, lactation stage and somatic cell count on milk yield and composition of Polish Black-and-White cows. *Anim. Sci.* Pap. Rep. **20:** 213–227 (2002).